Mechanisms of inotropic effects induced by nitric oxide

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(Ital Heart J 2001; 2 (Suppl 3): 48S-49S)

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Introduction

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Nitric oxide (NO) has been shown to modulate cardiac contractility. In physiologic concentrations (0.01-1 µM) NO induces a positive inotropic effect in different mammalian species including man. In contrast, concentrations of NO exceeding physiologic levels (> 1μ M) can reduce myocardial contractility. These effects of NO have been attributed to cGMP, the second messenger generated following NO-dependent activation of soluble guanylyl cyclase. Interestingly, it is not known whether endothelial cells or cardiomyocytes are the major cellular source of inotropic NO. Although the effects of NO on myocardial contraction are moderate, they might be important in regulation of myocardial function. Both impairment of endogenous NO production by endothelial NO-synthase and increased NO production by inducible NO-synthase could result in cardiodepressive effects.

Many years before organic nitrates were known to be NO donors, several laboratories reported positive inotropic effects of glyceryl trinitrate in animals and man¹⁻³. Most of the subsequent studies on the effects of exogenous NO have been performed with NO donors⁴. However, there are important differences between clinically used nitrovasodilators. This holds true for the mechanism and the kinetic of NO release and for the formation of toxic byproducts. For example, organic nitrates need enzymatic bioactivation to form NO, while sodium nitroprusside generates cyanide as cardiodepressive by-product⁴. The enzyme soluble guanylyl cyclase is the most important cellular target of NO (Fig. 1). This holds true for almost any cell type responding to NO⁵. In addition, NO has



Figure 1. Mechanism underlying the inotropic effects of nitric oxide (NO) on cardiac muscle. Activation of soluble guanylyl cyclase (sGC) and generation of cyclic guanosine monophosphate (cGMP) can induce inhibition of phosphodiesterase (PDE III) resulting in accumulation of cyclic adenosine monophosphate (cAMP) and activation of protein kinases (PKG) resulting in reduced transmembrane Ca^{2+} -current. Other mechanisms are the direct inhibition of myocardial adenosine triphosphate (ATP)-synthesis and of L-type Ca^{2+} -channels.

been shown to directly affect other cellular structures such as transmembrane ion channels and mitochondrial enzymes^{6,7}, al-though a cGMP-independent effect of NO on potassium channels has been questioned⁸. The known mechanisms underlying the inotropic effects of NO differ with respect to the concentration of NO and the expected effect on myocardial contractility.

cGMP-dependent mechanisms

The enzyme soluble guanylyl cyclase is found abundantly in vascular smooth muscle cells, neurons, platelets and also in human cardiomyocytes9,10. In mammalian cardiomyocytes the most important effector proteins of cGMP are the cGMP-dependent protein kinases and the cGMP-inhibited cGMP-phosphodiesterase¹¹. Stimulation of the protein kinases elicits inhibition of transmembrane Ca2+-current through voltage-dependent Ca²⁺-channels. This has been observed at high concentrations of NO (> 10 μ M) in different species such as frogs, guinea pigs and rats¹²⁻¹⁴. The underlying mechanism is not known but might involve phosphorylation of voltage-dependent Ca²⁺-channels¹⁵. The enzyme phosphodiesterase III hydrolyzes cAMP. It is important among phosphodiesterases in mammalian cardiomyocytes and inhibition of phosphodiesterase III has been observed at physiologic concentrations of NO donors (< 1 μ M)¹⁶. Binding of cGMP to phosphodiesterase III results in a reduced hydrolysis and accumulation of cAMP17. Studies with cardiomyocytes of frogs, guinea pigs and human atrial tissue showed that inhibition of phosphodiesterase by cGMP stimulates the transmembrane Ca²⁺-current^{9,14,18}.

cGMP-independent mechanisms

Studies with isolated dog ventricle showed inhibition of cellular respiration by the NO donor S-nitroso-N-acetylpenicillamine which was attributed to an impairment of mitochondrial electron transport⁷. Another mechanism which has been demonstrated in cells transfected with voltage-dependent Ca²⁺-channels is a cGMP-independent inhibition of transmembrane Ca²⁺current induced by high concentrations of NO donors (> 100 μ M)¹⁹.

References

- Raff WK, Drechsel U, Scholtholt J, Lochner W. Herzwirkung des Nitroglycerins. Pflugers Arch 1970; 317: 336-43.
- Strauer BE. Evidence for a positive inotropic effect of nitroglycerol on isolated human ventricular myocardium. Pharmacol Res Commun 1971; 3: 377-83.
- 3. Strauer BE, Scherpe A. Ventricular function and coronary hemodynamics after intravenous nitroglycerin in coronary artery disease. Am Heart J 1978; 95: 210-9.
- 4. Kojda G, Kottenberg K. Regulation of basal myocardial function by NO. Cardiovasc Res 1999; 41: 514-23.

- Lucas KA, Pitari GM, Kazerounian S, et al. Guanylyl cyclases and signaling by cyclic GMP. Pharmacol Rev 2000; 52: 375-414.
- Bolotina VM, Najibi S, Palacino JJ, Pagano PJ, Cohen RA. Nitric oxide directly activates calcium-dependent potassium channels in vascular smooth muscle. Nature 1994; 368: 850-3.
- Xie YW, Shen WQ, Zhao G, Xu XB, Wolin MS, Hintze TH. Role of endothelium-derived nitric oxide in the modulation of canine myocardial mitochondrial respiration in vitro. Implications for the development of heart failure. Circ Res 1996; 79: 381-7.
- Sausbier M, Schubert R, Voigt V, et al. Mechanisms of NO/cGMP-dependent vasorelaxation. Circ Res 2000; 87: 825-30.
- Kirstein M, Rivet-Bastide M, Hatem S, Bénardeau A, Mercadier JJ, Fischmeister R. Nitric oxide regulates the calcium current in isolated human atrial myocytes. J Clin Invest 1995; 95: 794-802.
- Lucas KA, Pitari GM, Kazerounian S, et al. Guanylyl cyclases and signaling by cyclic GMP. Pharmacol Rev 2000; 52: 375-414.
- Lohmann SM, Fischmeister R, Walter U. Signal transduction by cGMP in heart. Basic Res Cardiol 1991; 86: 503-14.
- Hartzell HC, Fischmeister R. Opposite effects of cAMP and cGMP on Ca²⁺-current in single heart cells. Nature 1986; 323: 273-5.
- Méry PF, Lohmann SM, Walter U, Fischmeister R. Ca²⁺ current is regulated by cGMP-dependent protein kinase in mammalian cardiac myocytes. Proc Natl Acad Sci USA 1991; 88: 1197-201.
- 14. Ono K, Trautwein W. Potentiation by cyclic GMP of β adrenergic effect on Ca²⁺ current in guinea-pig ventricular cells. J Physiol (Lond) 1991; 443: 387-404.
- 15. Jahn H, Nastainnczyk W, Röhrkasten A, Schneider T, Hofman F. Site-specific phosphorylation of the purified receptor for calcium-channel blockers by cAMP- and cGMP-dependent protein kinases, protein kinase C, calmodulin-dependent protein kinase II and casein kinase II. Eur J Biochem 1988; 178: 535-42.
- Weisshaar RE, Kobylarz-Singer DC, Kaplan HR. Subclasses of cyclic AMP phosphodiesterase in cardiac muscle. J Mol Cell Cardiol 1987; 19: 1025-36.
- Kojda G, Kottenberg K, Nix P, Schlüter KD, Piper HM, Noack E. Low increase in cGMP induced by organic nitrates and nitrovasodilators improves contractile response of rat ventricular myocytes. Circ Res 1996; 78: 91-101.
- Méry PF, Pavoine C, Belhassen L, Pecker F, Fischmeister R. Nitric oxide regulates cardiac Ca²⁺ current. Involvement of cGMP-inhibited and cGMP-stimulated phosphodiesterases through guanylyl cyclase activation. J Biol Chem 1993; 268: 26286-95.
- Hu H, Chiamvimonvat N, Yamagishi T, Marban E. Direct inhibition of expressed cardiac L-type Ca²⁺ channels by S-nitrosothiol nitric oxide donors. Circ Res 1997; 81: 742-52.